

# Nanowire platform for mapping neural circuits

Chong Xie and Yi Cui<sup>1</sup>

Department of Materials Science and Engineering, Stanford University, Stanford, CA 94305

**B**iological systems consist of a myriad of highly interactive and complex interconnections and pathways across multiple orders of length and time scales. Natural neuronal networks are valuable examples of such systems because they are important for understanding how the brain functions. Monitoring activities of a large number of neurons and intercommunication among them is critical for studying neuronal networks, for which passive microfabricated multielectrode arrays (MEAs) (1) and active planar silicon field effect transistor (FET) arrays (2) have been the two popular techniques. Now, a unique and more powerful technique has been developed, as demonstrated by Qing et al. in PNAS (3). The authors show that neuronal networks can now be studied with much higher spatial and temporal resolution while obtaining higher sensitivity of extracellular recording.

In the control center of the body, billions of neurons are linked together in a highly organized network for communication and control. A typical neuron consists of a cell body (soma), dendrites, and an axon (4). Neurons conduct information in two ways. One method is by electrical impulses traveling along the axon from one end of a neuron to the other; these impulses are known as action potentials. The second method is chemical transmission across the minute gap between neurons, the synapse, and is usually triggered by action potentials. Action potentials are based on excited movements of ions between the outside and inside of the cell membrane. Therefore, to study neuronal network behavior, the ability to monitor the electrochemical potential with high spatial and temporal resolution is essential.

Planar neuronal network systems, such as cultured neurons and brain slices, have been studied by MEAs (1). Extracellular field potentials could be recorded, but the spatial resolution is not enough to achieve single-cell level detection and signals are smaller than those detected by conventional micropipette electrodes. To overcome the problems associated with MEAs, planar FET arrays were developed. Because of the local amplification power of transistors, the weak signals do not have to travel through long leads and wires before being amplified (2). Thus signal amplitude and spatial resolution are greatly improved. However, because of the in-

trinsic size of the transistor and the limited capacitive coupling between the planar-gate electrode and neurons, it is necessary to have a fairly large gate-electrode area to obtain enough charge flow. This necessity limits further improvements of the spatial and temporal resolution of transistor arrays. In addition, a cleft filled by electrolyte is usually maintained between the neurons and transistors because of the flat substrate surface. This electrolyte layer causes a large leakage of potential signal. Therefore, a protruding device is preferred to achieve more efficient coupling.

Semiconductor nanowires (NWs) afford a unique, powerful chemical and biological sensing platform that has been developed in the past decade. Semiconductor NWs with diameters less than  $\sim 100$  nm have been synthesized with good control over composition, shape, and size (5–7). A variety of NW-based electronic devices (6), including FETs (8), have been fabricated, which have laid out a solid foundation for high performance sensors. Since the first demonstration that silicon NW FETs can be converted into sensors in 2001 (9), they have been shown to be able to detect charged chemicals, biomolecules, and viruses (9–15); they have also been used to study individual cultured neuron cells (16) and cardiac tissues (17). Nanowires have also been used to deliver biological molecules into cells (18, 19). In the present study (3), Qing et al. demonstrate that NW arrays on transparent substrates can be reliably used to study acute brain slices. The authors character-

ize the NW devices by simultaneously combining NW FET and patch clamp techniques to identify potential signals (Fig. 1). With NW devices, the authors not only identify action potential signals but also find additional features at earlier times. Using synaptic and ion-channel blockers during NW device sensing, the authors are able to assign these potential signals to presynaptic firing and postsynaptic depolarization. Interestingly, NW devices can access and identify different regions of neuron cells, which can either be close to somata or abundant in dendritic projections. With two-dimensional NW device array sensors interfacing with the olfactory cortex, the authors demonstrate multiplexed mapping and reveal spatially heterogeneous functional connectivity.

The silicon NW arrays demonstrated by Qing et al. afford unique features for neuronal network study (3). First, NW devices exhibit very high sensitivity and good signal-to-noise ratio. In the present study, the NW devices show signal amplitudes from 0.3 to 3.0 mV, which are higher than MEAs and planar FETs. Such high sensitivity results from the small diameter and protruding nature of NWs. NW devices protrude from

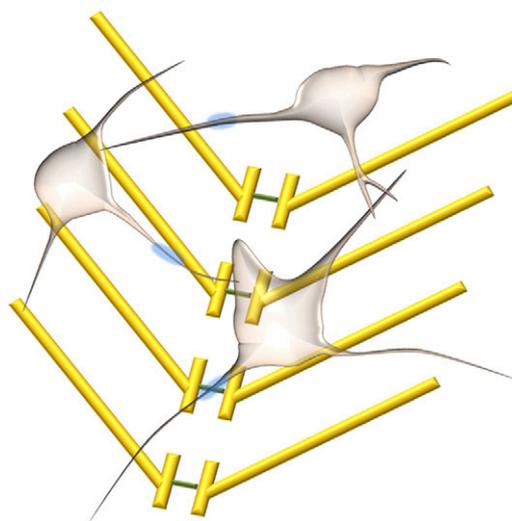


Fig. 1. Schematic of interconnected neurons and nanowire FET arrays.

Author contributions: C.X. and Y.C. wrote the paper.

The authors declare no conflict of interest.

See companion article on page 1882 in issue 5 of volume 107.

<sup>1</sup>To whom correspondence should be addressed. E-mail: yicui@stanford.edu.

